

Amino Acids Metabolism in the Muscle of Sheep fed with Mitchell Grass Hay Supplemented with *Gliricidia sepium*

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ABSTRACT

Leaf of *Gliricidia sepium* contains high amino acid that required for protein synthesis in the muscle. Supplementation of the legume leaves to low quality basal diet would improve amino acids amount to obtain an optimum growth of animal. The aim of experiment was to investigate the effect of *Gliricidia sepium* leaves supplementation to low quality basal diet on protein synthesis in muscle of animal. Eighteen sheep were divided into three groups based on live weight (27.6 ± 1.72 , 27.1 ± 2.26 and 27.5 ± 1.56 kg) of feed treatment, namely Mitchell grass hay (MG), *Gliricidia* (GS), and hay combined with *Gliricidia* (MGGS). All the diet was offered with the amount was determined to meet the maintenance metabolizable energy (ME) requirement of the animals. Feed utilization and amino acid metabolism in muscle of sheep was measured with radioactively labeled leucine, water and sodium bicarbonate. Results indicated that sheep in MG group had a negative energy balance, whereas sheep in MGGS group had the highest amount of ME available for growth. The rate of protein synthesis was higher in the MGGS group and tended to take up these amino acids in their hind-limb muscles, while animals in MG and GS groups tended to have net outputs of amino acids from their hind-limb muscles. Animals in the MGGS group synthesised more protein in their muscle than those in the GS and MG groups. It was concluded that the supplementation of *Gliricidia* at a ratio of 40:60 improved feed utilization of low quality basal diet. The improvement would be manifested in better digestible organic matter intake (DOMI) with subsequent better utilization of amino acids.

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INTRODUCTION

Supplementation of low quality basal diet (grass and agricultural by product) by leguminous leaves (*Leucaena keucocephala*, *Gliricidia sepium* and *Calliandra calothyrsus*, etc) aimed to improve ruminants productivity has been reported [1-4]. This supplementation was effectively increased the nutrient availability for the animals, in particular nitrogen required for optimum rumen microbial activity during rumen digestion of the feed. Improvement on feed intake, nutrient digestibility,

rumen fermentation efficiency and microbial protein as an effect of *Leucaena* or *Gliricidia* leaves supplementation has also been studied [5].

Animal's response to increasing nitrogen supply from legume leaves was manifested by improving in live weight gain. The intake and live weight gain of growth of Bali cattle fed with grass were observed when legume leaves was supplemented in the diet [6]. Previous study on the supplementation of *Gliricidia* leaves on grass basal diet indicated a significant increasing in dietary nitrogen retained in the sheep up to 18.4 % [7]. Dietary nitrogen is one of amino acids source for ruminant. Therefore, increasing in dietary retained nitrogen will be an indication of more supply of

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amino acids needed for protein synthesis. The important of dietary nitrogen for ruminants is not only to supply amino acids, but also as nitrogen sources for microbial protein synthesised in the rumen [8,9]. Rumen microbial protein is also a contributor of amino acids for ruminant animals. Thus information on the amino acids supply from *Gliricidia* leaves and its role in protein synthesis in the muscle of ruminant animal is required.

Protein as a source of amino acid is the most nutrient required for protein synthesis in the tissue muscle [10,11]. It was reported that large retention of nitrogen indicated a better utilization of dietary protein for muscle deposition, which in turn increase live weight gain [12]. Leucine and isoleucine were reported as essential amino acids required for protein synthesis. Among three legumes of *Leucaena*, *Gliricidia* and *Calliandra*, the *Gliricidia* and *Leucaena* contain higher amino acids compared to *Calliandra* leaves [13]. However, *Calliandra* was reported to contain high tannin as anti-nutritive that reduced the protein utilization by ruminant animals [14]. On the other hand, *Gliricidia* was reported produces higher leaves biomass per period of production compared to the two other legumes (*Leucaena* and *Calliandra*) [15]. Therefore the study was undertaken to determine the role of *Gliricidia* leaves as amino acids source for protein synthesis in sheep's muscle.

EXPERIMENTAL METHODS

Animals and feed

Eighteen sheep, approximately 18 months old, were used in a Randomized Block Design [16]. They were divided into three equal groups, evenly matched for live weight (LW). Each group was allocated at random to the following dietary treatments: Mitchell grass hay (*Astrelba lappacea*/MG), *Gliricidia* (*Gliricidia sepium*/GS), and MG combined with GS (MGGS). The mean \pm SE of initial LW of animals in each treatment group were 27.6 ± 1.72 (MG); 27.1 ± 2.26 (GS) and 27.5 ± 1.56 kg (MGGS).

The diets offered were designed to meet the Metabolize Energy (ME) for maintenance required by sheep according to National Research Council (NRC) [17]. The amount of *Gliricidia* leaves offered to animals in MGGS group was 37 % of total daily ration in dry matter (DM) base. This proportion was calculated to result in a nitrogen intake (NI):DOMI ratio of 0.04 [18] for optimum rumen microbial activity in feed digestion. Dry matter offered

was 815 g/day for MG and 578 g/day for GS, and 254 g/day of grass hay + 456 g/day of *Gliricidia* leaves for MGGS. The ration for each animal was divided into 12 equal portions, where each portion was given at 2-hour intervals from 07:00 AM to 15:00 PM. The balance of the 24 h ration was given to the animals at 17:00 PM. This feeding regimen was designed to provide a continuous flow of amino acids from the rumen into the duodenum, which should ensure a net amino acid absorption from the intestine [19].

Procedures

The experiment consisted of a 14-day period of adaptation to feeds and an 11-day of measurement period. The measurements undertaken were feed utilization (intake, digestibility and N balance) and the kinetics of urea, amino acid, and CO₂. Samples of feed offered, total feed residues and total faeces and urine excreted by the animals were collected each day for 7 days for feed utilization determination.

A set of chronic indwelling catheters was installed in the jugular veins, a femoral artery and a lateral saphenous vein of each sheep following procedure of Anisson [20] for determination of amino acids metabolism in the muscle. The procedure of catheters installation to the animals has been approved (Experimentation Ethics Approval No A635_01).

Infusate used in the experiment were L-[¹⁴C] leucine (13 kBq/mL), tritiated water [³HOH] (925 kBq/mL and 9.25 kBq/mL), and sodium [¹⁴C] bicarbonate (NaH¹⁴CO₃) (15.98 kBq/mL). The radioactivity of each infusate was determined immediately before and after the relevant infusion period. During the determination of amino acids metabolism, the L-[1-¹⁴C] leucine (0.33 μ Ci/mL) was infused continuously into the left jugular venous catheter of each experimental animal at the rate of 1 mL/min for 5.5 hours. It was assumed that the specific radioactivity (SRA) of leucine in the blood would start to plateau at two hours after the infusion started. Blood flow measurement was undertaken between the 3rd and 4th hour of L-[1-¹⁴C]leucine infusion [20] by using ³HOH (925 kBq/mL) solution that infused for 60 minutes into the right jugular venous catheter.

Two blood samples (5 mL and 10 mL each) were withdrawn from the femoral artery and lateral saphenous vein of each animal before and during the infusion of the leucine tracer for determination of radioactivity of leucine and CO₂. Five sets of three blood samples were withdrawn using syringes, at 20-minute intervals from the femoral artery and

lateral saphenous vein of each animal for determination of leucine radioactivity and amino acid concentration, determination of CO₂ radioactivity and blood gas determination.

The metabolized energy (ME) intake, ME required for maintenance and ME for growth were calculated based on the following equation [21]:

$$ME\text{-intake (MJ/d)} = [0.15x(DOMI/DMI)x100]xDMI,$$

$$ME\text{-maintenance (MJ/d)} = 1.2 + 0.13 LW$$

$$ME\text{-growth (MJ/d)} = ME\text{-i} - ME\text{-m}$$

Statistical analysis

All raw data were tabulated using Excel and analysed using IBM SPSS Statistics 20. Data were analyzed using ANOVA for the randomized block design. When significant effects of treatments were observed, differences among mean values were examined using Tukey's test [16].

RESULTS AND DISCUSSION

Feed utilization and energy balance

The grass and *Gliricidia* contain crude protein (5.7 % and 23 %); gross energy (18 and 17.26 MJ/kg DM), and NDF (55.3 and 51 %). Sheep in the MG and GS groups consumed similar amounts of DM but they consumed significantly less DM than the sheep in the MGGS group (Table 1). The apparent DM and organic matter (OM) digestibility values of the MGGS diet were between the values observed for MG and GS diet, with the lowest values recorded for the MG diet. Animals in GS group had the highest N intake, followed by MGGS and MG groups. About half of N consumed by animals in GS group was excreted as urinary N. The N value was significantly higher than those recorded in animals in the MGGS and MG groups. On the other hand, animals in the MG and MGGS groups excreted more than 50 % of their respective dietary N intake in faeces. The animals fed with MG had a negative N balance, while those fed with GS and MGGS had positive N balance values. The amount of N intake retained in animals in the MGGS group was significantly higher than that in animals in the GS group. Animals fed with MG was lost weight during the observation period, while animals fed with GS and MGGS gained LW. The LW gain was significantly higher in MGGS group. The feed conversion ratio was lower in MGGS than in GS group. The value of NI:DOMI ratio for the MGGS group was between the values recorded for the other two groups; the highest being observed in the GS group.

Table 1. Feed utilization by sheep fed Mitchell grass hay supplemented or not with *Gliricidia*.

Variable	Treatment			SE
	MG	GS	MGGS	
Intake (g/d)				
DM	577 ^b	608 ^b	746 ^a	60
OM	511	569	612	71
App. Digestibility (%)				
DM	44 ^c	55 ^a	49 ^b	1.4
OM	52 ^c	59 ^a	56 ^b	0.8
N balance (g/d)				
N intake	3.4 ^c	15.7 ^a	10 ^b	0.8
N faeces	2.6 ^c	7.0 ^a	5.2 ^b	0.6
N urine	0.9 ^c	7.9 ^a	2.9 ^b	0.4
N retained	-0.12 ^c	0.80 ^b	1.84 ^a	0.7
NI : DOMI	0.01 ^c	0.05 ^a	0.03 ^b	0.33
LW gain LWG (g)	-90 ^c	28 ^b	80 ^a	53
FC ratio(gDMI/gLWG)	-	22 ^b	9 ^a	8.9

Within rows, mean with different superscript different significantly
 App. Digestibility = apparent digestibility; DM = dry matter; OM = organic matter; N = nitrogen; NI = nitrogen intake; DOMI = digestible organic matter intake; LW = live weight; Fcratio = Feed conversion ratio; MG = Mitchell grass; GS = *Gliricidia sepium*; MGGS = Mitchell grass+*Gliricidia sepium*.

All animals required similar amounts of ME for maintenance. Animals in the MGGS group had the highest ME intake, while animals in the MG and GS groups consumed similar amounts of ME (Table 2). Sheep fed MG had a negative energy balance, while sheep fed MGGS had the highest amount of ME available for growth.

Table 2. Energy utilization by sheep fed with Mitchell grass hay supplemented with *Gliricidia*

Variables	Treatments			± SE
	MG	GS	MGGS	
Energy balance (MJ/d)				
ME-i ¹	4.0 ^b	5.0 ^b	5.7 ^a	0.46
ME-m ²	4.7	4.7	4.8	0.34
ME-g ³	-0.7 ^b	0.3 ^a	0.9 ^a	0.35
ME-retained (%ME-i)	0 ^c	6.0 ^b	15.8 ^a	2.24

Within rows, means with different superscript, differ significantly.
 ME-I = Metabolize energy intake; ME-m = Metabolize energy maintenance; ME-g = metabolize energy growth

Urea kinetics

The urea kinetics in sheep fed the experimental feed measured during the experiment is presented in Tabel 3.

Sheep fed with GS had the highest plasma urea concentration, urea entry rate (UER), amount of urea excreted through urine and volume of excreted urine among the treatment groups. The lowest corresponding values were observed in the MG fed animals. The urinary N present as urea N (%) was similar for the GS and MGGS sheep, both values were higher than that observed in the MG fed animals.

Table 3. Urea kinetics in sheep fed with Mitchell grass hay supplemented with Gliricidia

Variables	Treatments			± SE
	MG	GS	MGGS	
Plasma urea concentration (mg/100mL)	27 ^c	80 ^a	55 ^b	6
Urea entry rate (UER) (g/d)	27 ^c	82 ^a	58 ^b	7
Urinary urea (g/d)	0.8 ^b	10.3 ^a	4.3 ^b	4.8
Urinary N as urea-N (%)	42 ^b	61 ^a	70 ^a	8
Urine output (mL/d)	928 ^b	1645 ^a	953 ^b	402

Within rows, values with different superscript differ significantly
 UER was estimated using: $Y = 0.7932 X + 16.027$,
 Y = urea entry rate (g/d); X = plasma urea concentration (mg/100 mL)

Protein synthesis

Based on the kinetics of ³HOH solution that was infused during the 60 minutes period into the right jugular venous of the sheep, the blood flow can be determined. Thus, with the combination of L-[1-¹⁴C] leucine infusion, the metabolism of leucine in the whole body of the sheep can be calculated.

Dietary treatments significantly influenced leucine entry rate (LER) (Table 4). The LER in MGGS groups was the highest compared to other groups. It was not significantly different with the LER observed in the GS group, but was significantly different with the LER in MG group.

Dietary treatments significantly affected the proportion of decarboxylated leucine. Animals fed with GS had the highest percentage of decarboxylated leucine. This also was shown by the amount of CO₂ produced from leucine. However, the percentage of decarboxylation and CO₂ amount produced from leucine in GS group were not different with the values observed in MGGS group. These values were significantly higher than in MG animals. The estimated mean rate of protein synthesis was higher in MGGS group and significantly higher than that observed in MG group, but did not differ significantly from GS group.

Table 4. Leucine metabolism in the whole body of sheep fed with Mitchell grass hay supplemented with Gliricidia

Variables	Treatments			± SE
	MG	GS	MGGS	
<i>Whole body¹⁾</i>				
LER (mmol/h)	13.6 ^b	17.6 ^{ab}	20.4 ^a	2.3
CO ₂ leucine(mmol/h)	0.4 ^b	0.6 ^a	0.6 ^{ab}	0.1
Leucine DO (%)	9 ^b	42 ^a	24 ^{ab}	11
Protein synthesised ²⁾ (g/d)	404 ^b	520 ^{ab}	606 ^a	69

Within rows, values with different superscripts differ significantly
 LER = Leucine entry rate; Leucine DO = Leucine decarboxylase
 1) It was assumed that empirical factor for respiratory output of mature sheep is 0.235 mmol CO₂/min/kg LW.
 2) It was assumed that leucine content in sheep muscle is 127.6 mmol/kg muscle [18].

The dietary treatments did not have a significant effect on the extraction of leucine by the hind-limb muscle (Table 5). However, treatments did have a significant effect on the proportion of leucine taken up by muscle that was catabolised. The animals in GS group had the highest proportion of catabolised leucine while those in MG group had the lowest. The contribution of leucine to muscle CO₂, likewise, was significantly affected by the dietary treatments and was highest in GS group and lowest in MG group. The amounts of leucine involved in muscle protein synthesis were higher in GS and MGGS groups than in MG group. Animals in MGGS group synthesized more protein in their muscle than those animals in GS and MG groups. Dietary treatments had no significant effect on arterial concentrations of essential amino acid (Table 5), although the concentrations tended to be higher in GS group than MG and MGGS groups.

Table 5. Leucine kinetics and protein synthesis in total muscle of sheep fed Mitchell grass hay supplemented or not with Gliricidia

Variables	Treatments			± SE
	MG	GS	MGGS	
<i>Hind-limb muscle:</i>				
Leucine fraction extracted (%)	19	20	17	4
True leucine extracted (µM)	28	34	29	7
Leucine catabolism (% of uptake)	17 ^c	58 ^a	36 ^b	8
leucine to muscle CO ₂ (%)	0.1 ^b	0.4 ^a	0.3 ^b	0.13
Leucine for protein synthesis ¹⁾ (µmole/h/kg muscle)	164 ^b	328 ^a	310 ^a	79
<i>Total muscle:</i>				
CO ₂ from leucine (mmole/h)	0.04 ^c	0.25 ^a	0.25 ^b	0.06
Protein synthesised ²⁾ (g/d)	33 ^b	34 ^b	63 ^a	11
Protein synthesis (muscle/whole body,%)	9	7	11	

Within rows, values with different superscripts differ significantly
 1) It was assumed that muscle mass is 25 % of LW.
 2) With the assumption that protein fraction in muscle of sheep six months of age was 0.16 [18]

Data on the arterial concentration of Isoleucine and Leucine in sheep fed experimental feed measured during the experiment is presented in Table 6.

Table 6. Concentrations of leucine and isoleucine in arterial sheep (µM) fed with Mitchell grass hay supplemented with Gliricidia

Variables	Treatments			±SE
	MG	GS	MGGS	
<i>Arterial concentration (µM)</i>				
Isoleucine	79	93	84	4
Leucine	150	192	163	8

The amount of grass fed to the animals were calculated to provide 4.8 MJ/day of ME required for maintenance of sheep. However, animals in the MG

group did not eat all of the diet offered. Consequently, the animal in this group was in negative energy balance (Table 2) and lost of their LW (Table 1). Animal in GS and MGGS groups consumed slightly higher ME (6 % and 19 % respectively) than other group. As a consequence, both groups had some ME available for growth (Table 2) and thus gained LW. These are consistent with the observation that both groups also had positive N balance (Table 1). It is clear that the loss weight of animals in MG group was due to the low DMI and DM digestibility of the grass (44 %; Table 1) as well as low protein content of MG (5.7 %). The result was similar with the other report that showed low digestibility (46 %) of the grass with low CP content (5.6 %) [22]. The addition of GS to MG in MGGS group increased intake and digestibility by 29 % and 11 %, respectively. Such improvements are in agreement with other research that found that the addition of GS to Buffel grass improved digestibility [23].

The most probably reason of DMI and digestibility improvement in MGGS group was due to the shift in NI:DOMI ratio of 0.01 in MG group to 0.03 in MGGS group when GS was supplemented. The finding was coherent with the study indicated that rumen microbial activity improved when grass supplemented with GS [22]. Moreover, supplementation of *Gliricidia* improved digestibility of Buffel grass by hairy sheep [23]. The improvement in feed efficiency in MGGS group was shown by the lower feed conversion ratio (Table 1). The MGGS group consumed 9 g of DM/unit of LWG compared with 22 g DM/unit LWG consumed by GS group. This improvement was most likely due to better nutrient balance.

Negative N balance observed in MG group was consistent with the value of amino acid outflow from hind-limb muscle (Table 5). Urea concentration in plasma was positively associated with N intake, with reflects the catabolism of amino acids, both within the rumen and at the body tissue level ($Y = 4.01 X + 15.045$, $R^2 = 0.8049$; where Y is the plasma urea concentration and X is the nitrogen intake). Differences in the rates of urine secreted by the sheep fed with the different diets probably were due largely to differences in urea concentration in the plasma. The large urine output in GS group reflected the increased osmotic load in blood of animals caused by high urea concentration in plasma (Table 3).

The increasing LER in MG group, through GS to the MGGS group, most probably reflected the increasing amounts of leucine (and other amino acids) absorbed from the alimentary tract of the animals in the respective groups (Table 5). It is interesting that although the GS group

consumed highest amount of N (Table 1), the LER for GS and MGGS groups was similar although the MGGS group consumed less N. It might caused by more balanced NI:DOMI ratio in MGGS, where there was a greater amount of microbial amino acids available for absorption from the small intestine. The apparent increase in LER in GS group would be due to high amount of leucine supplied by *Gliricidia* (Table 5), as also reported that *Gliricidia* is one of legumes leaves that contains high leucine and isoleucine [13].

There was a positive relationship between CP intake and plasma concentration of essential amino acids. This again lends further support to the suggestion that there was increased flow of amino acids from the intestine to the bloodstream of animals with increased N intake. The concentration of leucine in plasma of MGGS group appear to be lower than that in GS group, although the LER in MGGS group was higher than that in GS group. A high LER would reflect a high utilization rate of leucine, it might be suggested that the apparently lower plasma concentration of leucine in the MGGS group reflected the higher utilization rate of the amino acid in protein synthesis in these animals.

Although higher proportions of the leucine taken up by the hind-limb muscle of animals in GS and MGGS groups that were catabolised, the amounts of amino acids available in hind limb for protein synthesis was higher than that in hind limb of the MG group when blood flow rates across the hind-limb muscle beds (Table 5) are taken into account.

No significant dietary treatment effects were observed on the A-V concentration difference of leucine and isoleucine across the hind-limb muscles (Table 5 and 6). This suggests that probably, the dietary treatments did not affect the transporters of the two amino acids into muscle cells of the animals under observation. Overall, the A-V concentration difference of amino acids across the hind-limb muscles tended to be generally positive in the MGGS group and generally negative in the MG and GS groups (Table 5). It is most likely that the positive A-V concentration difference of leucine and isoleucine (Table 5 and 6) in hind-limb muscles of MGGS sheep indicate significant increase in the rate of protein synthesis and net protein deposition in the muscles. This is consistent with the result of N retained (Table 1). The finding of study also consistent with the report that the amount of nitrogen retained would reflect the amount of protein synthesised in muscle [12].

CONCLUSION

Supplementation of *Gliricidia* on Mitchell grass hay increased dietary retained energy (18.9 %

of energy intake) and protein synthesis due to increasing on supply of amino acids from *Gliricidia* as well as balance of nitrogen and energy availability as indicated by ratio of N:DOMI (0.04). These results was supported by significantly higher weight gain of the animals after supplementation of *Gliricidia* leaves.

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REFERENCES

1. B.O. Oduguwa, A.O. Oni and O.M. Arigbede, *Trop. Anim. Health Prod.* **45** (2013) 1363. DOI: 10.1007/s11250-013-0370-y
2. T. Panjaitan, M. Fauzan and Dahlanuddin et al., *Tropical Grasslands-Forrajes Tropicales* **2** (2014) 116. DOI: 10.17138/TGFT(2)116-118
3. S. Gunasekaran, K. Viswanathan and C. Bandeswaran, *International Journal of Science Environment and Technology* **3** (2014) 1767.
4. N. Hilmiati, Sutartha and T. Panjaitan, *International Journal of Sustainable Development* **09** (2016) 47.
5. M. Wanapat, K. Sungchhang and P. Sineenart, *Journal of Animal Science and Biotechnology* **4** (2013) 32. <https://doi.org/10.1186/2049-1891-4-32>
6. Dahlanuddin, O. Yanuario and D.P. Poppi, *Animal Production Science* **54** (2013) 915.
7. Y. Widiawati, E. Teleni and Suharyono, *Atom Indonesia* **40** (2014) 121.
8. M.A. Brooks, R.M. Harvey and N.F. Johnson, *Journal of Animal Science* **90** (2012) 4985.
9. J.N. Kim, E.D. Henriksen and I.K. Cann, *Appl Applied and Environmental Microbiology* **80** (2014) 3095. <http://www.springerplus.com/content/2/1/483>.
10. S. Francisco, D. Pacheco and H. Blair, *Springplus.* **2** (2013) 438.
11. Guoyao Wu, W.B. Fuller and Zhaolai Dai, *Annual Review of Animal Biosciences* **2** (2014) 569.
12. E.M. Alves, D.R. Magalhães and M.A. Freitas, *Acta Scientiarum Animal Science* **36** (2014) 55.
13. P.A. Aye and M.K. Adegun, *Biology Journal of North America* **4** (2013) 71.
14. S.J. Bunglavan and N. Dutta, *Journal of Livestock Science* **4** (2013) 67.
15. Steven Franzel, Sammy Carsan and Ben Lukuyu, *Current Opinion in Environmental Sustainability* **6** (2014) 98.
16. W.W. Daniel, *Biostatistics: A Foundation Analysis in the Health Science*, 5th ed., John Wiley and Sons. Inc, USA (1991) 207.
17. Anonymous, *Nutrient Requirements of Small Ruminants: Sheep, Goats, Cervids, and New World Camelids*, Animal Nutrition Series, National Research Council (2007) 07.
18. T.J. Hackmann and J.L. Firkins, *Front Microbiology* **15** (2015) 465.
19. I.D. Hume, D.R. Jacobson and J.R. Mitchell, *Journal of Nutrition* **102** (1972) 495.
20. E. Teleni and E.F. Annison, *Australian Journal of Biology Science* **39** (1986) 271. DOI.org/10.1071/B19860271
21. Anonymous, *Energy Allowances and Feeding Systems for Ruminants*, Great Britain, Department of Agriculture and Fisheries for Scotland, Northern Ireland, Department of Agriculture, London (1975) 15.
22. A.T.W. Raharjo, W. Suryapratama and T. Widyastuti, *Journal of Animal Science* **1** (2013) 796. (In Indonesian)
23. J.N. Avilés-Nieto, J.L. Valle-Cerdán and Castrejón-Pineda, *Tropical Animal Health and Production* **45** (2013) 1357.